

INSET PROGRAM

# 2D DNA FILM

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## What They Are Doing

- Construction of 2D and 3D nanonetworks from relatively large pieces of DNA
- Used for construction of nanorobotic devices, nanosensors, and analysis of nanomaterials

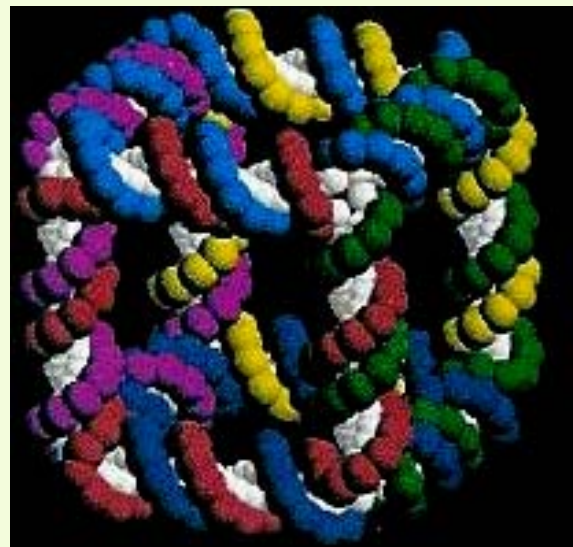
## What We Are Doing

- Using small, single-stranded pieces of DNA called oligonucleotides
- Facilitate faster self-assembly and allow smaller features
- Creating a film or ribbon of DNA for same applications

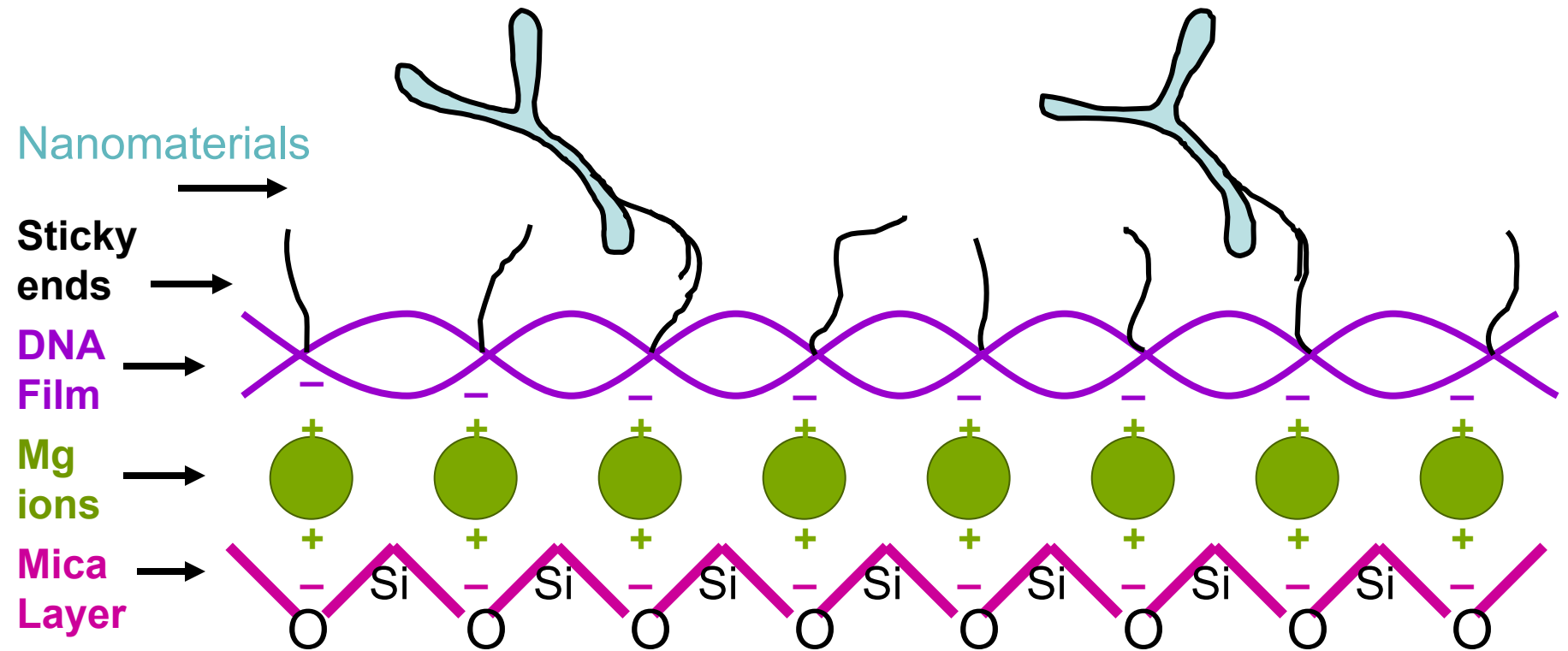
Truncated Octahedron



Cube



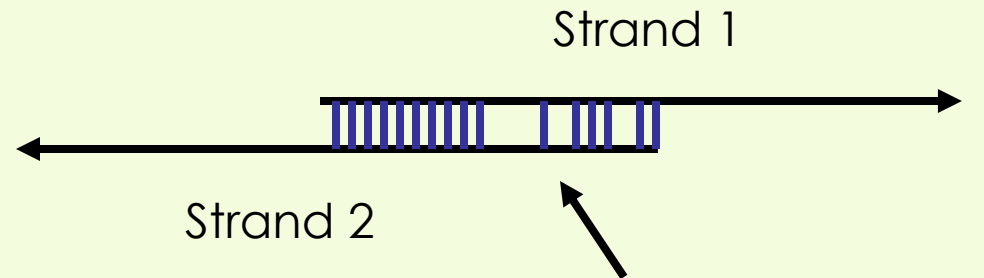
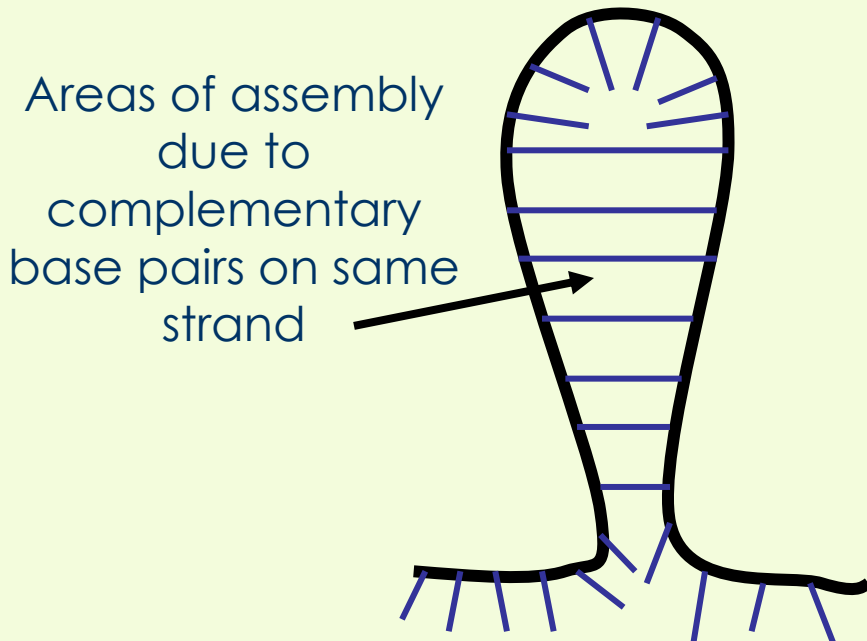
# Molecular View



# Designing the Oligonucleotides

Hairpins: DNA bases on one strand bind to itself

Dimers: bases from two strands of DNA bind at unfavorable locations

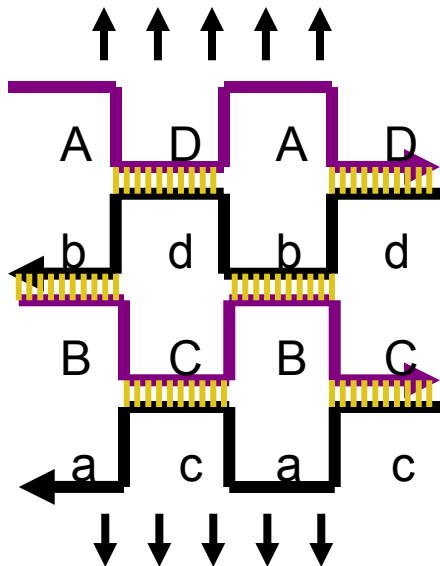
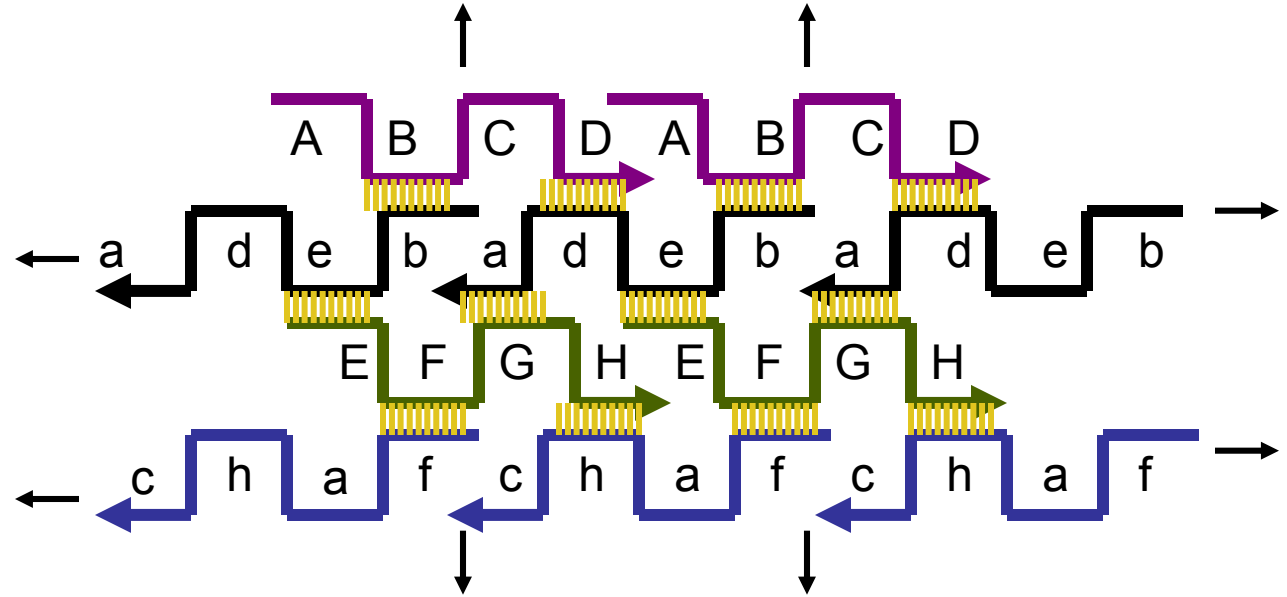


Areas of assembly due to complementary base pairs at various sites on separate strands of DNA

These cause interruptions or defects in the film formations

# The Difference Between DNA Film and DNA Ribbons

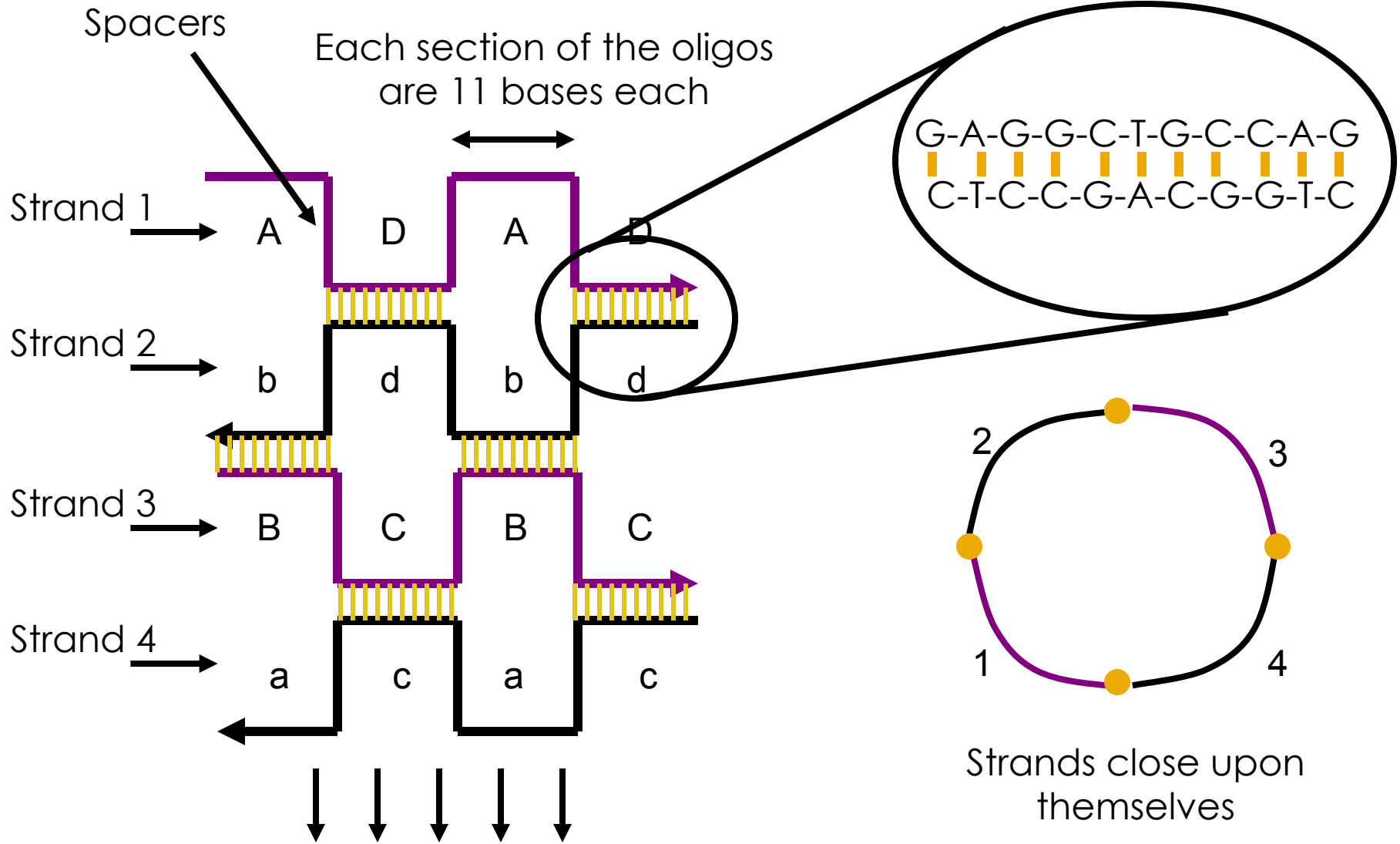
DNA Film=  
has growth in all  
four directions



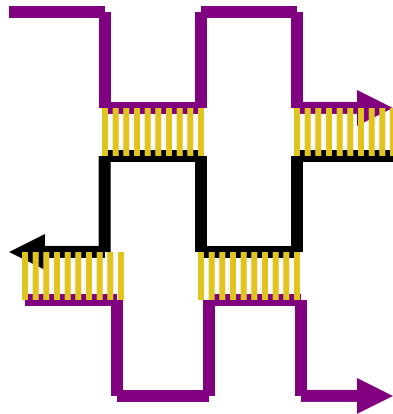
DNA Ribbon=  
only has growth in  
two directions

What forms, film or  
ribbons, all  
depend upon the  
sequence of the  
base pairs on the  
oligonucleotides

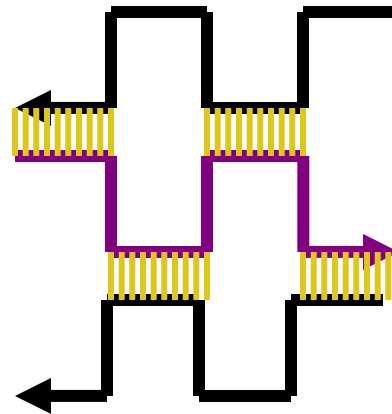
# 2D Film Assembly



# Assembly of 2D DNA Ribbons with “Triplet” Technique

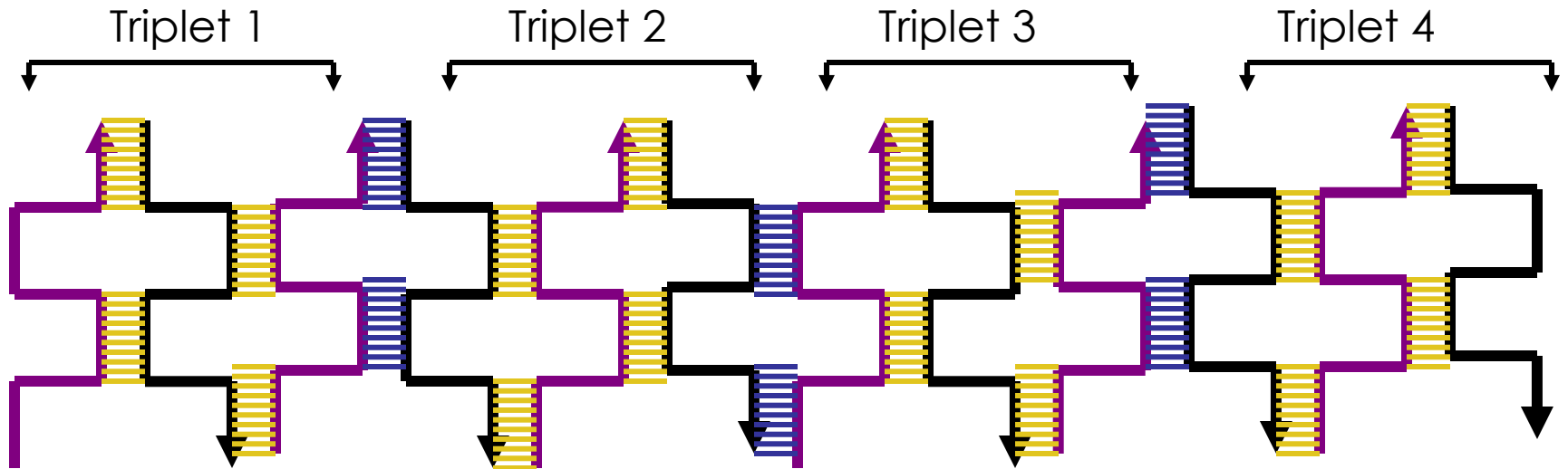


Triplet 1



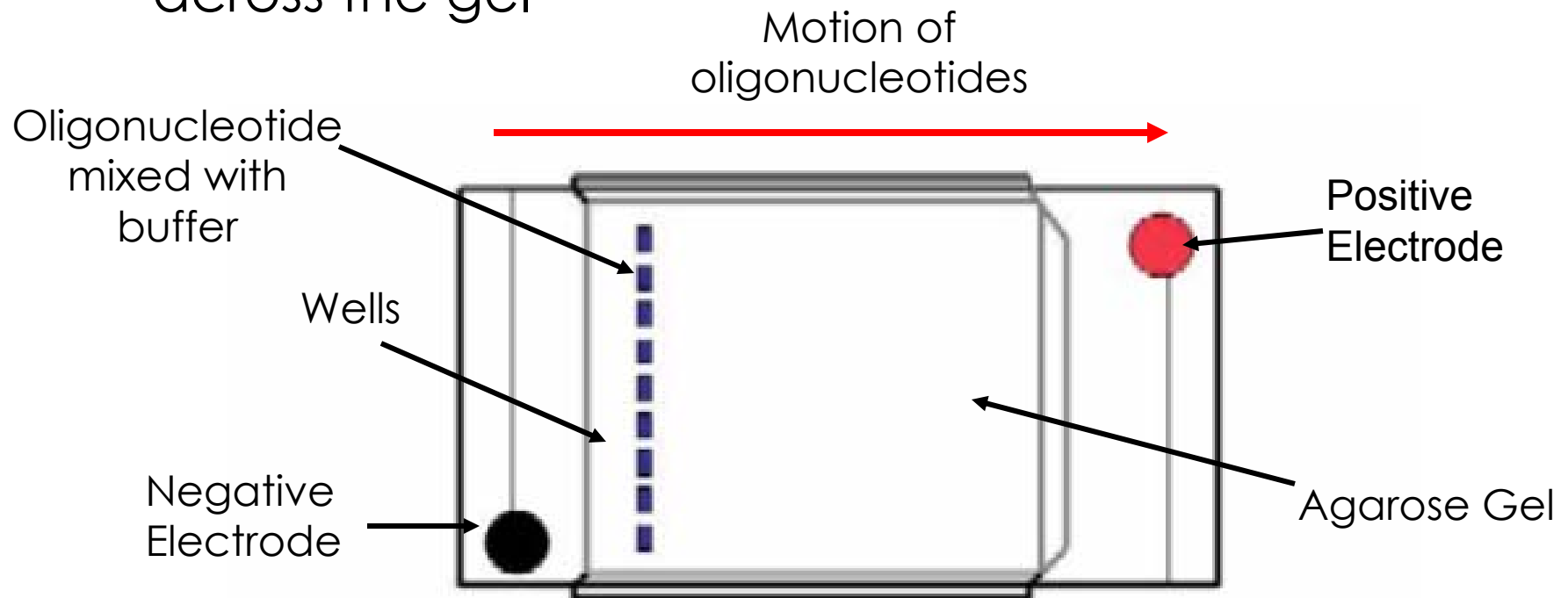
Triplet 2

Assemble Triplets Separately



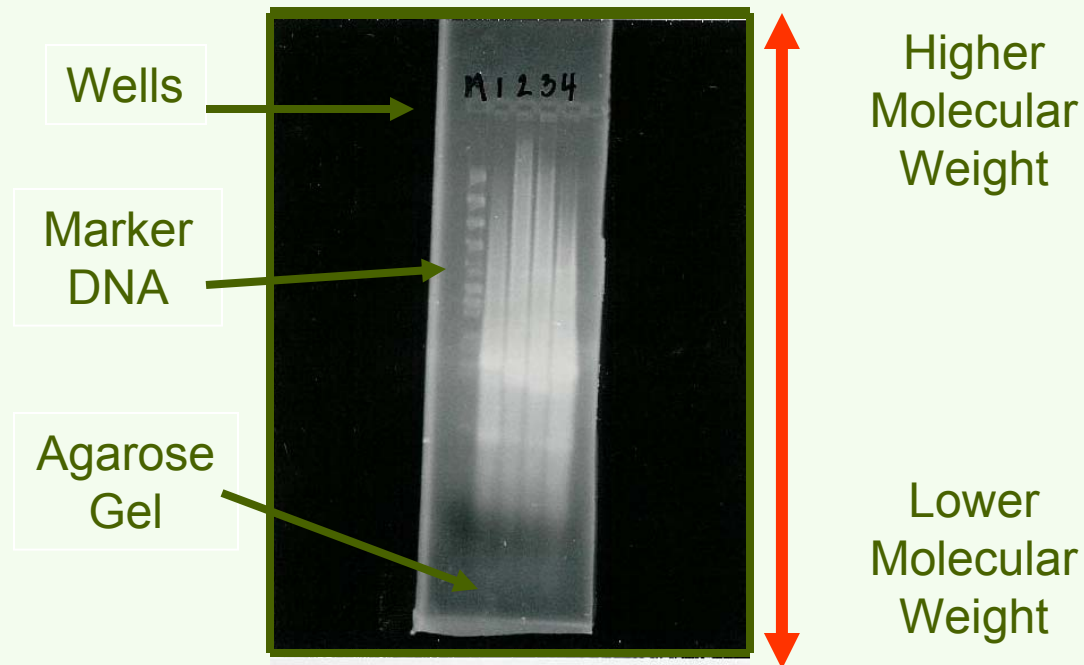
# Agarose Gel Electrophoresis

- Will separate the structures that formed according to size
- Higher molecular weight structures stay closer to the wells
- Lower molecular weight structures migrate across the gel



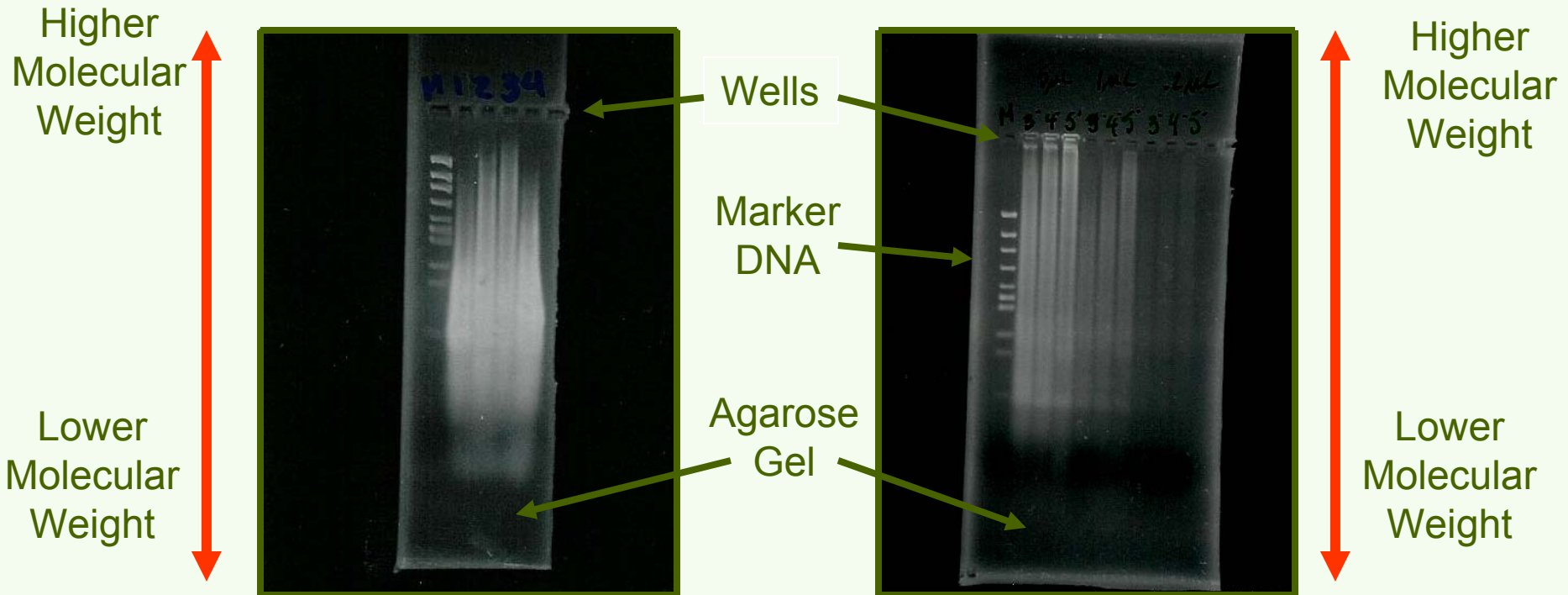


# Oligonucleotide Assembly: Without Triplet Technique



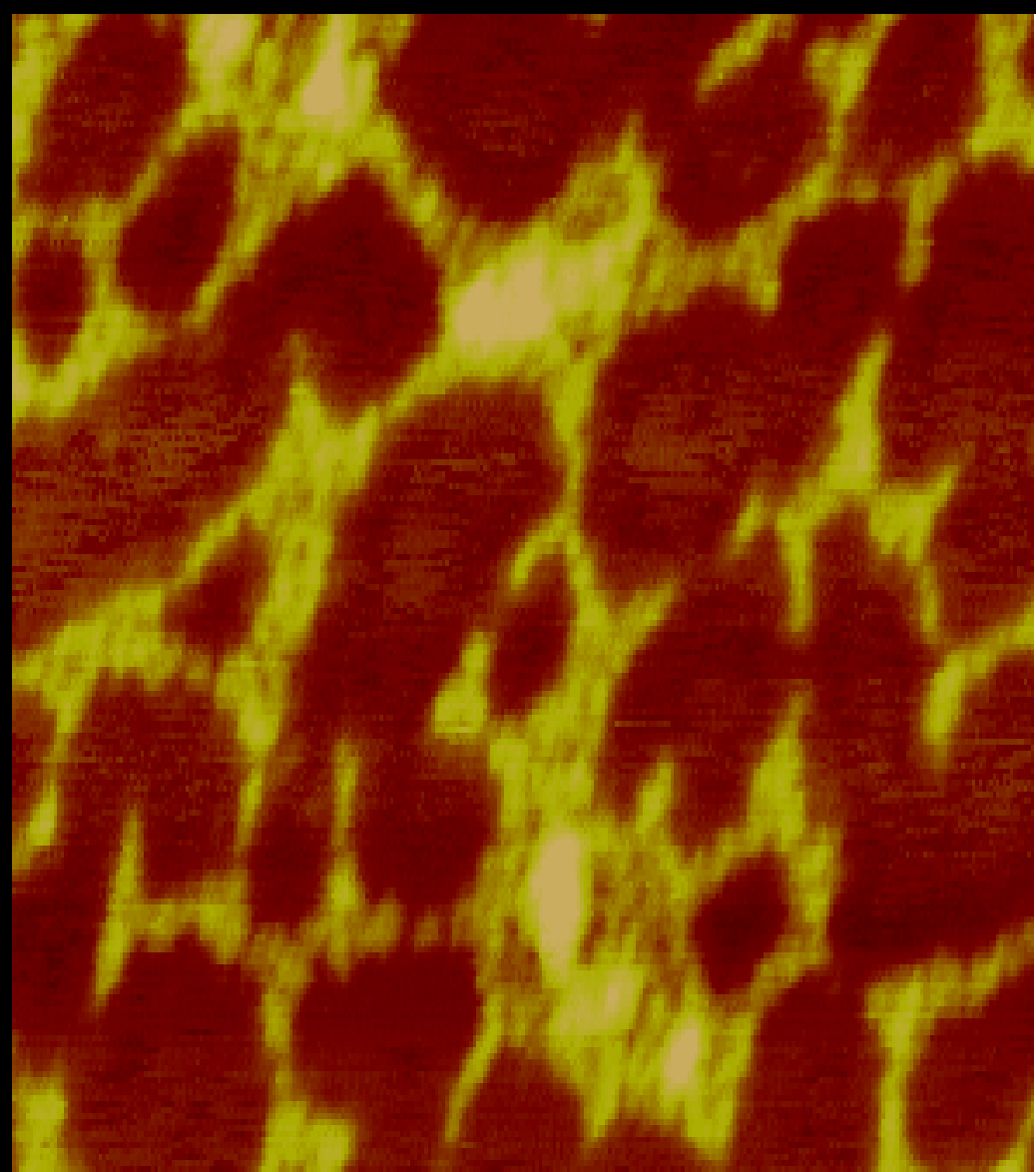
Mixtures prepared with four types of oligonucleotide strands per sample. Concentration of magnesium chloride varied for samples 1-4. Not many high molecular weight compounds formed.

# Using Oligonucleotides Triplets: to Prevent DNA Bundles



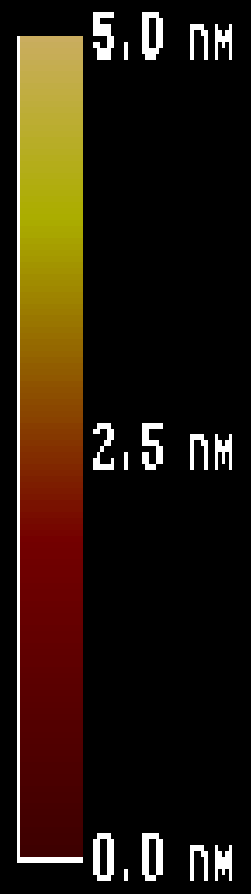
Picture 1: Oligonucleotides assembled with three strands per solution- triplets 1,2,3 & 4.

Picture 2: 10 microL of each of the four triplets mixed together three times to assemble film-heated at 37°, 47°, & 57°C.



750

500

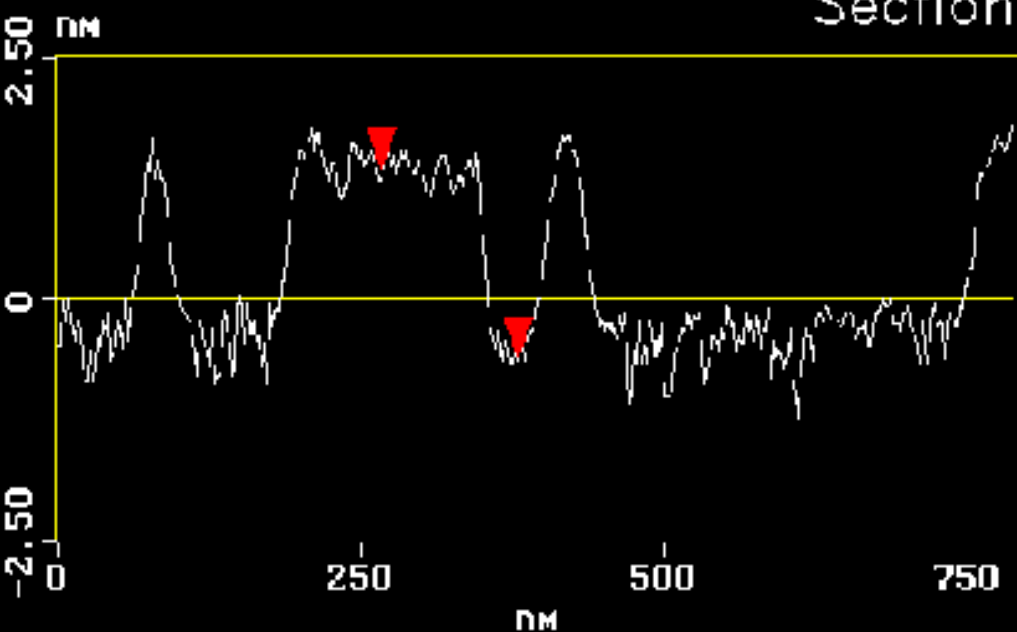


5.0 nm

2.5 nm

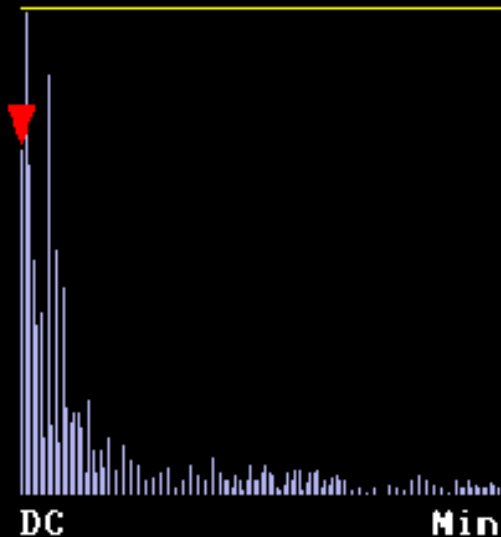
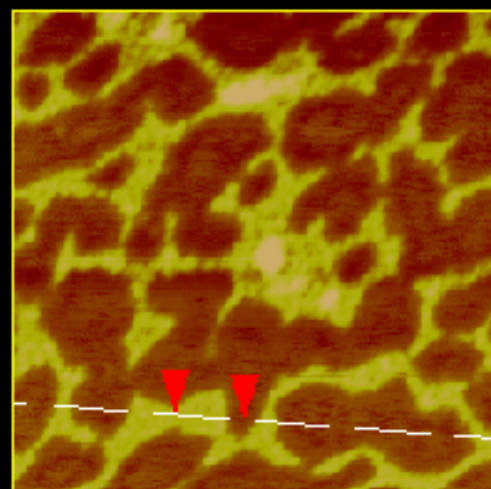
0.0 nm

# Section Analysis



L	112.72 nm
RMS	0.790 nm
lc	DC
Ra(lc)	0.400 nm
Rmax	1.705 nm
Rz	1.196 nm
Rz Cnt	4
Radius	802.95 nm
Sigma	0.388 nm

## Spectrum



Surface distance	113.11 nm
Horiz distance(L)	112.72 nm
Vert distance	1.950 nm
Angle	0.991 °
Surface distance	
Horiz distance	
Vert distance	
Angle	
Surface distance	
Horiz distance	
Vert distance	
Angle	
Spectral period	DC
Spectral freq	0 Hz
Spectral RMS amp	0.0006 nm

07011606.002

## In the Immediate Future

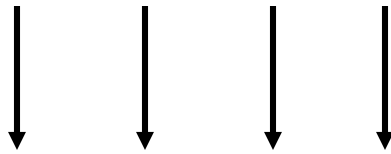
- Design a new set of oligonucleotides that are more specific in their interactions
- Assemble the oligonucleotides on a mica crystal surface

## Acknowledgements

- Helen Hansma and Emin Oroudjev
- INSET

# To Prevent Hairpins and Dimers

Desired interactions have  
highest melting point usually  
between 46°C and 58°C

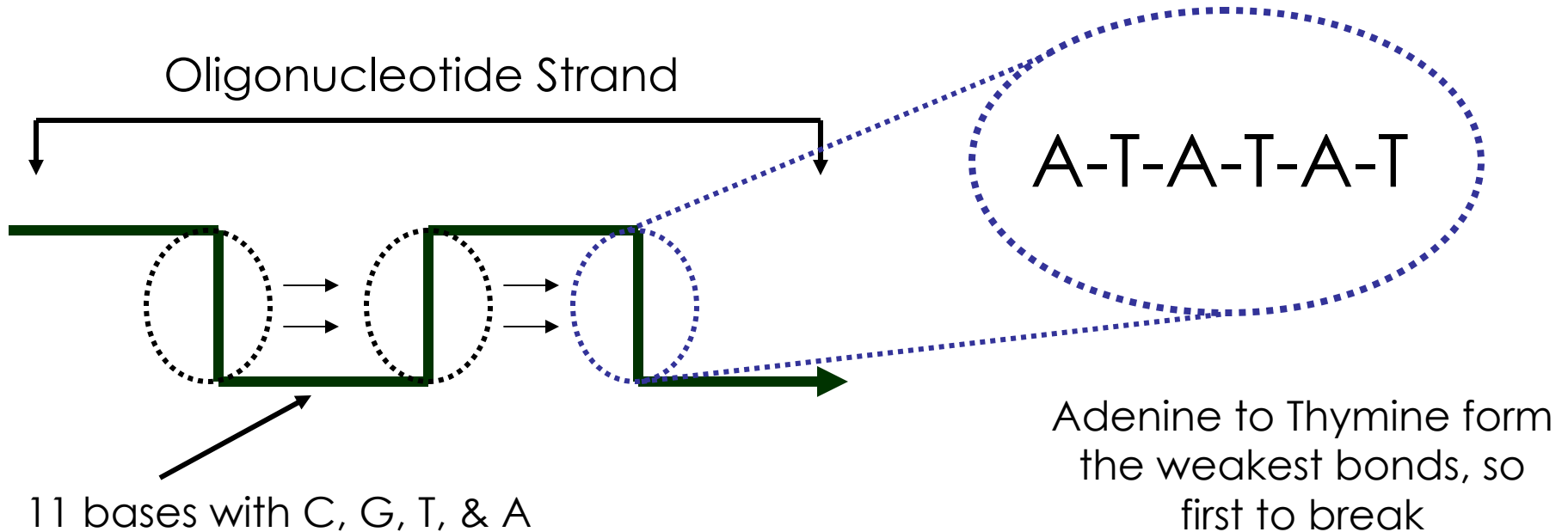


Hairpin and dimer  
formations have a  
melting point between  
36°C and 42°C

Heat the oligonucleotides up to 98°C and let cool for an hour and a half. This allows the correct interactions between the strands to occur while the hairpin and dimer interactions form and then melt again.

# The Importance of Spacers

Oligonucleotide Strand



- How long should they be?
- What bases should they be composed of?
- Should they interact with a spacer on a different strand?

# Agarose Gel Electrophoresis

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